

Short Communication



# Hub Genes and Therapeutic Pathways of CTRP9 in Cardiac Ischemia/Reperfusion Injury: A Bioinformatics Perspective

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## Abstract

**Background:** Ischemia/reperfusion injury (IRI) is a significant contributor to cardiac morbidity, while therapeutic options remain limited. According to previous evidence, C1q/TNF-related protein 9 (CTRP9), an adipokine with cardiometabolic regulatory properties, may serve as a potential modulator of myocardial injury. In this study, comprehensive bioinformatics analyses were primarily employed to identify hub genes and elucidate key therapeutic pathways associated with CTRP9 in the context of IRI.

**Methods:** An interrelation analysis of hub genes was conducted to identify direct and indirect interactions. The resulting network and enrichment data were extracted from Cytoscape-GeneMANIA3.6.0, based on a genome-wide human interaction map, for further analysis and visualization.

**Results:** Protein-protein interaction networks and functional enrichment analyses revealed that CTRP9 exerts cardioprotective effects primarily through the activation of adenosine monophosphate-activated protein kinase (AMPK) signaling, which leads to reduced cardiomyocyte apoptosis and enhanced angiogenesis. Notably, bioinformatics data suggested several downstream effectors, such as *ucp1*, *gsk3b*, and *rps6kb1*, as well as collagen-related genes (i.e., *col4a2*, *col14a1*, and *col18a1*), linked to the beneficial effects of CTRP9, considering it a promising therapeutic option related to IRI.

**Conclusion:** These findings recommend several hub genes and pathways that may serve as novel therapeutic targets, highlighting the potential of CTRP9-based interventions for managing IRI-induced cardiac damage and improving clinical outcomes, particularly in cases of myocardial damage caused by IRI. Uncovering all possible underlying mechanisms could enhance our ability to better address the pathological sequelae following IRI.

**Keywords:** AMPK signaling, Angiogenesis, C1q/TNF-related protein 9 (CTRP9), Ischemia/Reperfusion injury, Bioinformatics analyses

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## Introduction

Restoring blood flow, also known as reperfusion, to ischemic tissue results in adverse events, such as endothelial cell injury, local and systemic inflammation, tissue edema, and increased vascular permeability.<sup>1,2</sup> This phenomenon, characterized as ischemic/reperfusion injury (IRI), can diminish the efficacy of reperfusion therapy, which is the mainstay of managing ischemic conditions.<sup>3,4</sup> Notably, our recent review dominantly elucidated the potential role of C1q tumor necrosis factor-related protein-9 (CTRP9) in favor of IRI prevention and treatment.<sup>5</sup>

The CTRP9 is one of the members of the adiponectin

paralog family, drawing attention for its potential roles, especially in cardiovascular physiopathology. Initially identified as a cardiokine, CTRP9 is abundantly expressed in cardiac tissue, where it functions with significant autocrine and paracrine effects. Emerging evidence highlights CTRP9's protective role against myocardial IRI, a major contributor to cardiac dysfunction following acute myocardial infarction.<sup>6</sup> During IRI, the sudden restoration of blood flow leads to oxidative stress, inflammation, and cardiomyocyte apoptosis, exacerbating myocardial damage. CTRP9 has been shown to mitigate these deleterious effects by activating intracellular signaling pathways that promote cell survival



and inhibit apoptosis. In this regard, experimental studies also demonstrated that CTRP9 deficiency exacerbates cardiac injury and dysfunction post-IRI, whereas CTRP9 overexpression or supplementation confers significant protection, highlighting its therapeutic potential.

One critical and well-established pathway implicated in CTRP9-mediated cardioprotection is the adenosine monophosphate-activated protein kinase (AMPK) signaling cascade.<sup>7</sup> AMPK serves as a cellular energy sensor and regulator, orchestrating metabolic homeostasis and stress responses.<sup>8,9</sup> Furthermore, CTRP9-induced AMPK activation inhibits apoptosis and endoplasmic reticulum stress in cardiomyocytes subjected to ischemic insult, thereby preserving myocardial function.

To the best of our knowledge, CTRP9 activates AMPK signaling directly through binding to the adiponectin receptor 1 (AdipoR1), but not AdipoR2, on target cells such as endothelial cells and cardiomyocytes, leading to the phosphorylation and activation of AMPK, and subsequent downstream protective cellular responses, e.g., nitric oxide production through the AMPK-endothelial nitric oxide synthase pathway.<sup>9</sup> During the AdipoR1-AMPK axis, CTRP9 stimulates the phosphorylation of AMPK at the Thr172 site on the  $\alpha$  subunit.<sup>10,11</sup>

Beyond the AMPK signaling pathway, CTRP9 also engages other pro-survival signaling pathways, including protein kinase A-cAMP response element-binding protein axis, which further contributes to its anti-apoptotic effects in cardiomyocytes.<sup>12</sup> The interaction between CTRP9 and molecular chaperones, such as calreticulin, facilitates the activation of these protective pathways, highlighting a complex regulatory network. Additionally, CTRP9 influences vascular remodeling and angiogenesis, which are essential processes for myocardial repair and recovery after ischemic injury.

Considering the indispensable role of CTRP9 in alleviating IRI, the identification of involved effectors and relevant signaling pathways must be addressed. Accordingly, this study aims to provide insights into potential strategies for enhancing cardiac repair potential against ischemic stress. It particularly explores the potency of candidate factors using the bioinformatics analysis regarding the interplay of *c1qtnf9* with possible signaling pathways.

## Methods

An interrelation analysis of the identified hub genes to determine their interactions was performed, essentially among the genes that interacted with one another directly or indirectly. Using Cytoscape-GeneMANIA (version 3.6.0),<sup>13</sup> it was attempted to predict gene function and construct gene interaction networks, integrating numerous biological data types, such as protein-protein interactions, genetic interactions, co-expression, co-localization, and shared protein domains, to identify genes related to a given query gene set, *c1qtnf9*. The final network and enrichment results were exported from

Cytoscape-GeneMANIA for downstream analysis, to interpret and visualize the significant gene sets identified, and figure preparation according to the results obtained from a genome-wide map of human genome interaction. This is a comprehensive, global representation of how various regions of the genome physically associate and communicate within the three-dimensional space of the cell nucleus, offering an essential understanding of gene regulation mechanisms that extend beyond the simple linear DNA sequence.

## Results and Discussion

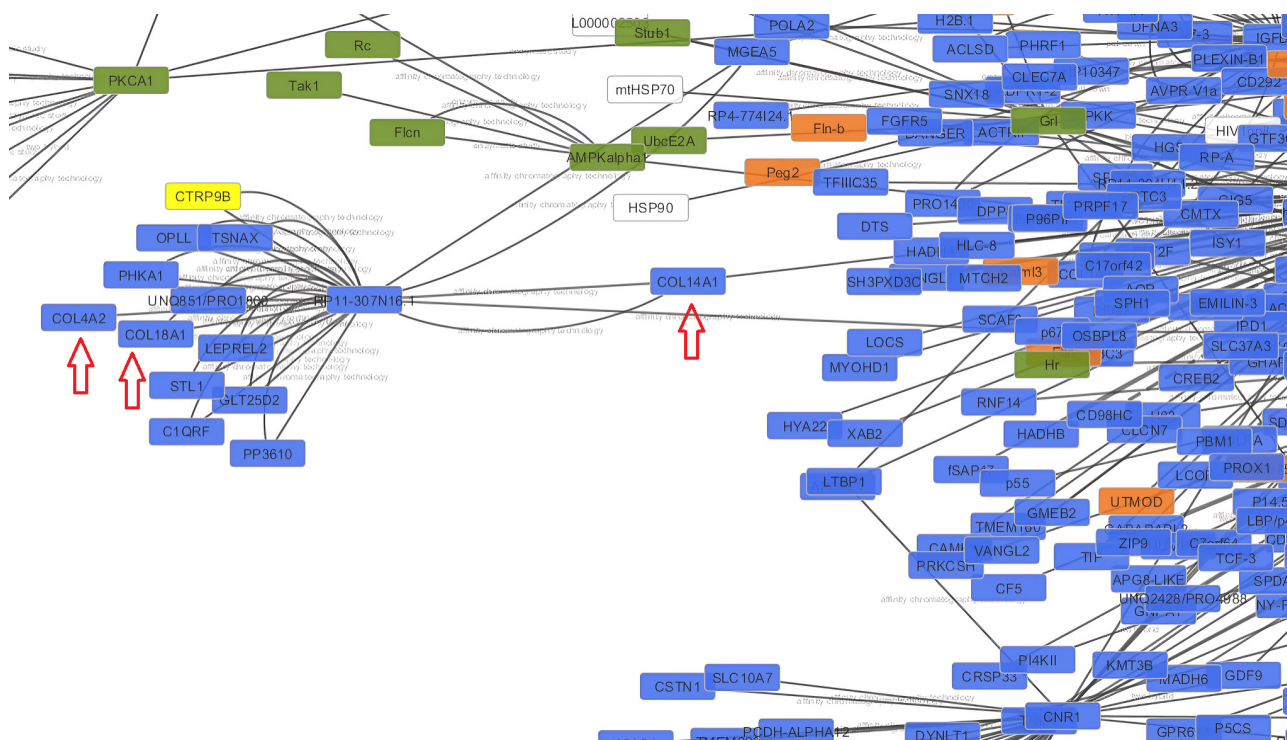
Notably, our results revealed that the CTRP-9-related encoded gene (*c1qtnf9*) has the potential to interact with three genes, including *rps6kb1*, *gsk3b*, and *ucp1* (Figure 1). The *rps6kb1* gene encodes ribosomal protein S6 kinase beta-1 (RPS6KB1), a serine/threonine kinase in humans, which has a crucial role in protein synthesis<sup>14</sup> and belongs to the PI3K-dependent signaling pathway. Mechanistically, the *rps6kb1* is mainly activated by the phosphoinositide 3-kinase/mammalian target of rapamycin (PI3K/mTOR) pathway, regulating protein synthesis via ribosomal protein S6 phosphorylation.<sup>15</sup> Previous results elegantly demonstrated the close interaction of the *rps6kb1* factor with the mTOR/autophagy pathway (as AMPK downstream), in which the binding of *rps6kb1* to mTOR-related effectors promotes the autophagic response.<sup>16</sup> In ischemic cardiac microvascular endothelial cells, it has also been shown that the inhibition of *rps6kb1* promotes the down-regulation of vascular endothelial growth factor via the hypoxia-inducible factor 1- $\alpha$ -related pathway and aberrant angiogenesis by suppressing MAPK/PI3K/Akt signaling, further highlighting its importance in vascular repair during ischemia.<sup>17</sup>

*Gsk3b*, the second identified factor by bioinformatics analysis, is an isoform of the GSK-3 enzyme, known as an energy regulatory sensor, mainly acting through Wnt/ $\beta$ -catenin and PI3K/AKT pathways. The GSK3b has been a promising therapeutic target for some ischemic diseases.<sup>18,19</sup> However, a dual function of GSK3 $\beta$  has been identified in myocardial injury caused by IRI. During extended ischemia, the activation of GSK3 $\beta$  stimulates autophagy by inhibiting mTOR, which promotes the survival of cardiomyocytes. Conversely, in the reperfusion phase, inhibiting GSK3 $\beta$  helps protect the heart by reducing excessive autophagy, thereby mitigating myocardial damage.<sup>20</sup> Hence, it is evident that the opposite regulation of GSK3 $\beta$  throughout IRI plays a crucial role in balancing cell survival and death in ischemic heart disease. Collectively, both *gsk3b* and *rps6kb1* have a cross-link in mTOR signaling,<sup>21</sup> influencing both angiogenesis and autophagy during ischemia and modulating signaling cascades affecting vascular function.

The latter factor, *Ucp1* gene, encoding a transmembrane protein, UCP1, also named thermogenin, is crucial in energy-induced hemostasis in epicardial adipose tissue and can be altered during cardiometabolic syndromes.<sup>22-24</sup>







**Figure 2.** Utilizing a Comprehensive Gene-Pathway Database to Uncover Shared Pathways Influenced by CTRP9: Evaluation of Direct Interactions Between the CTRP9 gene (*c1qtnf2*) and Related Genes by Cytoscape  
 Note. CTRP9: C1q tumor necrosis factor-related protein-9

integrity and angiogenesis, which are a part of multi-locus genetic risk scores for coronary artery diseases, increasing hemorrhagic stroke risk. These traits can affect atherosclerotic plaque stability, preventing macrovascular pathologies and the risk of MI.<sup>27</sup> A preprint study also revealed that dysregulation in collagen-related pathways has a crucial role in the abolished hearts and identified molecular defects using single-cell mRNA sequencing data in the subset of newborns. Noteworthy, some pre-collagen pathways, including MAS and connective tissue growth factor (MAS-CTGF-collagen pathway), may also act to modulate pro-collagen expression, such as *Col1A1*, *Col1A2*, *Col3A1*, and *Col4A2*.<sup>28</sup> Moreover, it has been well-documented that the mutations in *col4a1/a2* genes potentially disrupt basement membranes, enhancing the risk for small vessel disease, including deep intracerebral hemorrhage,<sup>29</sup> hypertension, aortic dilation, and coronary artery dissections.<sup>30, 31</sup>

Furthermore, compelling evidence demonstrated the key role of collagen XIV, especially *Col14a1*, in the regulation of early stages of fibrillogenesis, cardiomyocyte proliferation, cell survival, and heart maturity, which was further confirmed by hub-gene assessment.<sup>32, 33</sup>

Regarding the *Col18a1*, it has also been proven that by engaging a structural role in the basement membrane alongside its angiostatic fragment endostatin, *Col18a1* is considered a critical regulator of vascular homeostasis in CVD.<sup>34</sup> In this line, preclinical data declared that *Col18a1* deficiency contributes to atherosclerosis progression, microvascular damage,<sup>35</sup> and cardiac remodeling.<sup>36</sup> These findings further indicate that *Col18a1* is a potential

biomarker and therapeutic target in various cardiovascular conditions.

## Conclusion

These results highlight key hub genes and pathways that could serve as potential therapeutic targets, proposing the promise of CTRP9-based treatments more likely mediated by *ucp1*, *gsk3b*, and *rps6kb1*, as well as collagen-related genes, for addressing myocardial injury due to IRI and improving patient outcomes. Uncovering the full spectrum of underlying mechanisms may further enhance our ability to effectively manage the pathological consequences following IRI.

## Ethical statement

Not applicable.

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This study received no financial support.

## Conflict of interests declaration

The authors declare no conflict of interests.

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## Data availability statement

The corresponding author will provide the datasets used and/or analyzed during the current work upon reasonable request.

**Author contributions****Conceptualization:** Aysa Rezabakhsh.**Investigation:** Seyyed-Reza Sadat-Ebrahimi, Masoud Khalifezadeh.**Methodology:** Shahrouz Ghaderi.**Software:** Shahrouz Ghaderi.**Supervision:** Aysa Rezabakhsh.**Writing—original draft:** Seyyed-Reza Sadat-Ebrahimi.**Writing—review & editing:** Aysa Rezabakhsh.**Consent for publication**

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**References**

- Valikesserlis I, Athanasiou AA, Stakos D. Cellular mechanisms and pathways in myocardial reperfusion injury. *Coron Artery Dis.* 2021;32(6):567-77. doi: [10.1097/mca.0000000000000997](#).
- Cowled P, Fitridge R. Pathophysiology of reperfusion injury. In: Fitridge R, ed. *Mechanisms of Vascular Disease: A Textbook for Vascular Specialists*. Cham: Springer International Publishing; 2020. p. 415-40. doi: [10.1007/978-3-030-43683-4\\_18](#).
- Hassankhani H, Soheili A, Shams Vahdati S, Mozaffari FA, Fraser JF, Gilani N. Treatment delays for patients with acute ischemic stroke in an Iranian emergency department: a retrospective chart review. *Ann Emerg Med.* 2019;73(2):118-29. doi: [10.1016/j.annemergmed.2018.08.435](#).
- Rahimi Mamaghani A, Abdi F, Abbasalizad Farhangi M. Food insecurity prevalence and its demographic, anthropometric, and nutritional determinants among overweight and obese patients with diabetes and coronary artery disease. *Int J Drug Res Clin.* 2024;2:e15. doi: [10.34172/ijdrcl.2024.e15](#).
- Sadat-Ebrahimi SR, Amini H, Rahbarghazi R, Habibollahi P, Ghaderi S, Rajabi H, et al. Putative therapeutic impacts of cardiac CTRP9 in ischaemia/reperfusion injury. *J Cell Mol Med.* 2022;26(11):3120-32. doi: [10.1111/jcmm.17355](#).
- Sun Y, Yi W, Yuan Y, Lau WB, Yi D, Wang X, et al. C1q/tumor necrosis factor-related protein-9, a novel adipocyte-derived cytokine, attenuates adverse remodeling in the ischemic mouse heart via protein kinase A activation. *Circulation.* 2013;128(11 Suppl 1):S113-20. doi: [10.1161/circulationaha.112.000010](#).
- Kambara T, Shibata R, Ohashi K, Matsuo K, Hiramatsu-Ito M, Enomoto T, et al. C1q/tumor necrosis factor-related protein-9 protects against acute myocardial injury through an adiponectin receptor I-AMPK-dependent mechanism. *Mol Cell Biol.* 2015;35(12):2173-85. doi: [10.1128/mcb.01518-14](#).
- Marino A, Hausenloy DJ, Andreadou I, Horman S, Bertrand L, Beauloye C. AMP-activated protein kinase: a remarkable contributor to preserve a healthy heart against ROS injury. *Free Radic Biol Med.* 2021;166:238-54. doi: [10.1016/j.freeradbiomed.2021.02.047](#).
- Hu Q, Qu W, Zhang T, Feng J, Dong X, Nie R, et al. C1q/Tumor necrosis factor-related protein-9 is a novel vasculoprotective cytokine that restores high glucose-suppressed endothelial progenitor cell functions by activating the endothelial nitric oxide synthase. *J Am Heart Assoc.* 2024;13(4):e030054. doi: [10.1161/jaha.123.030054](#).
- Zheng Q, Yuan Y, Yi W, Lau WB, Wang Y, Wang X, et al. C1q/TNF-related proteins, a family of novel adipokines, induce vascular relaxation through the adiponectin receptor-1/AMPK/eNOS/nitric oxide signaling pathway. *Arterioscler Thromb Vasc Biol.* 2011;31(11):2616-23. doi: [10.1161/atvbaha.111.231050](#).
- Li L, Gu Z, Zhang J. CTRP9 overexpression attenuates palmitic acid-induced inflammation, apoptosis and impaired migration in HTR8/SVneo cells through AMPK/SREBP1c signaling. *Exp Ther Med.* 2022;24(1):459. doi: [10.3892/etm.2022.11386](#).
- Zhao D, Feng P, Sun Y, Qin Z, Zhang Z, Tan Y, et al. Cardiac-derived CTRP9 protects against myocardial ischemia/reperfusion injury via calreticulin-dependent inhibition of apoptosis. *Cell Death Dis.* 2018;9(7):723. doi: [10.1038/s41419-018-0726-3](#).
- Lin A, Wang RT, Ahn S, Park CC, Smith DJ. A genome-wide map of human genetic interactions inferred from radiation hybrid genotypes. *Genome Res.* 2010;20(8):1122-32. doi: [10.1101/gr.104216.109](#).
- Bdzhola A, Malanchuk O, Palchevsky S, Gout I, Filonenko V, Zhyvoloup A. Co-expression of the RPS6KB1 and PDPK1 genes for production of activated p70S6K1 using bac-to-bac baculovirus expression system. *Mol Biol Rep.* 2025;52(1):130. doi: [10.1007/s11033-024-10136-0](#).
- Bahrami F, Ataie-Kachoei P, Pourgholami MH, Morris DL. p70 Ribosomal protein S6 kinase (Rps6kb1): an update. *J Clin Pathol.* 2014;67(12):1019-25. doi: [10.1136/jclinpath-2014-202560](#).
- Datan E, Shirazian A, Benjamin S, Matassov D, Tinari A, Malorni W, et al. mTOR/p70S6K signaling distinguishes routine, maintenance-level autophagy from autophagic cell death during influenza A infection. *Virology.* 2014;452-453:175-90. doi: [10.1016/j.virol.2014.01.008](#).
- Dai GH, Ma PZ, Song XB, Liu N, Zhang T, Wu B. MicroRNA-223-3p inhibits the angiogenesis of ischemic cardiac microvascular endothelial cells via affecting RPS6KB1/hif-1a signal pathway. *PLoS One.* 2014;9(10):e108468. doi: [10.1371/journal.pone.0108468](#).
- Wang W, Li M, Wang Y, Wang Z, Zhang W, Guan F, et al. GSK-3 $\beta$  as a target for protection against transient cerebral ischemia. *Int J Med Sci.* 2017;14(4):333-9. doi: [10.7150/ijms.17514](#).
- Shams Vahdati S. Anti-platelets and antithrombotic co-ingestion in prevention of recurrent stroke. *Int J Drug Res Clin.* 2025;3(1):e4. doi: [10.34172/ijdrcl.2025.e4](#).
- Zhai P, Sciarretta S, Galeotti J, Volpe M, Sadoshima J. Differential roles of GSK-3 $\beta$  during myocardial ischemia and ischemia/reperfusion. *Circ Res.* 2011;109(5):502-11. doi: [10.1161/circresaha.111.249532](#).
- Hermida MA, Dinesh Kumar J, Leslie NR. GSK3 and its interactions with the PI3K/AKT/mTOR signalling network. *Adv Biol Regul.* 2017;65:5-15. doi: [10.1016/j.jbior.2017.06.003](#).
- Kalinovich AV, de Jong JM, Cannon B, Nedergaard J. UCP1 in adipose tissues: two steps to full browning. *Biochimie.* 2017;134:127-37. doi: [10.1016/j.biochi.2017.01.007](#).
- Flouris AD, Shidlovskii YV, Shaposhnikov AV, Yepiskoposyan L, Nadolnik L, Karabon L, et al. Role of UCP1 gene variants in interethnic differences in the development of cardio-metabolic diseases. *Front Genet.* 2017;8:7. doi: [10.3389/fgene.2017.00007](#).
- Chechi K, Vijay J, Voisine P, Mathieu P, Bossé Y, Tchernof A, et al. UCP1 expression-associated gene signatures of human epicardial adipose tissue. *JCI Insight.* 2019;4(8):e123618. doi: [10.1172/jci.insight.123618](#).
- Pravednikova AE, Shevchenko SY, Kerchev VV, Skhirtladze MR, Larina SN, Kachaev ZM, et al. Association of uncoupling protein (UCP) gene polymorphisms with cardiometabolic diseases. *Mol Med.* 2020;26(1):51. doi: [10.1186/s10020-020-00180-4](#).
- Deng Y, He Y, Xu J, He H, Zhang M, Li G. Cardiac fibroblasts regulate myocardium and coronary vasculature development via the collagen signaling pathway. *bioRxiv [Preprint]*. September 12, 2024. Available from: <https://www.biorxiv.org/content/10.1101/2024.09.11.612512v1>.
- Yang W, Ng FL, Chan K, Pu X, Poston RN, Ren M, et al. Coronary-heart-disease-associated genetic variant at the COL4A1/COL4A2 locus affects COL4A1/COL4A2 expression, vascular cell survival, atherosclerotic plaque stability and risk of myocardial infarction. *PLoS Genet.* 2016;12(7):e1006127.

- doi: [10.1371/journal.pgen.1006127](https://doi.org/10.1371/journal.pgen.1006127).
28. Chatterjee A, Barnard J, Moravec C, Desnoyer R, Tirupula K, Karnik SS. Connective tissue growth factor dependent collagen gene expression induced by MAS agonist AR234960 in human cardiac fibroblasts. *PLoS One*. 2017;12(12):e0190217. doi: [10.1371/journal.pone.0190217](https://doi.org/10.1371/journal.pone.0190217).
29. Rannikmäe K, Davies G, Thomson PA, Bevan S, Devan WJ, Falcone GJ, et al. Common variation in COL4A1/COL4A2 is associated with sporadic cerebral small vessel disease. *Neurology*. 2015;84(9):918-26. doi: [10.1212/wnl.0000000000001309](https://doi.org/10.1212/wnl.0000000000001309).
30. Steffensen LB, Rasmussen LM. A role for collagen type IV in cardiovascular disease? *Am J Physiol Heart Circ Physiol*. 2018;315(3):H610-25. doi: [10.1152/ajpheart.00070.2018](https://doi.org/10.1152/ajpheart.00070.2018).
31. Gasparini S, Balestrini S, Saccaro LF, Bacci G, Panichella G, Montomoli M, et al. Multiorgan manifestations of COL4A1 and COL4A2 variants and proposal for a clinical management protocol. *Am J Med Genet C Semin Med Genet*. 2024;196(4):e32099. doi: [10.1002/ajmg.c.32099](https://doi.org/10.1002/ajmg.c.32099).
32. Tao G, Levay AK, Peacock JD, Huk DJ, Both SN, Purcell NH, et al. Collagen XIV is important for growth and structural integrity of the myocardium. *J Mol Cell Cardiol*. 2012;53(5):626-38. doi: [10.1016/j.yjmcc.2012.08.002](https://doi.org/10.1016/j.yjmcc.2012.08.002).
33. Ma Z, Wang X, Lv Q, Gong Y, Xia M, Zhuang L, et al. Identification of underlying hub genes associated with hypertrophic cardiomyopathy by integrated bioinformatics analysis. *Pharmgenomics Pers Med*. 2021;14:823-37. doi: [10.2147/pgpm.S314880](https://doi.org/10.2147/pgpm.S314880).
34. Ambade AS, Naranjo M, Tuhy T, Yu R, Marimoutou M, Everett AD, et al. Collagen 18A1/endostatin expression in the progression of right ventricular remodeling and dysfunction in pulmonary arterial hypertension. *Am J Respir Cell Mol Biol*. 2024;71(3):343-55. doi: [10.1165/rcmb.2024-0039OC](https://doi.org/10.1165/rcmb.2024-0039OC).
35. Khoshneviszadeh M, Henneicke S, Pirici D, Senthilnathan A, Morton L, Arndt P, et al. Microvascular damage, neuroinflammation and extracellular matrix remodeling in Col18a1 knockout mice as a model for early cerebral small vessel disease. *Matrix Biol*. 2024;128:39-64. doi: [10.1016/j.matbio.2024.02.007](https://doi.org/10.1016/j.matbio.2024.02.007).
36. Moulton KS, Olsen BR, Sonn S, Fukui N, Zurakowski D, Zeng X. Loss of collagen XVIII enhances neovascularization and vascular permeability in atherosclerosis. *Circulation*. 2004;110(10):1330-6. doi: [10.1161/01.Cir.0000140720.79015.3c](https://doi.org/10.1161/01.Cir.0000140720.79015.3c).